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Title: Uniparental chicken offsprings derived from oogenesis of chicken primordial germ cells (ZZ)

Short title: Oogenesis of primordial germ cells (ZZ) produces uniparental chicken offspring

Summary sentence:

Cultured chicken primordial germ cells (ZZ) possess the developmental plasticity to differentiate into functional ova in the ovary of germline chimeric chicken host, and give rise to uniparental chicken offspring.

Keywords: gametogenesis, sexual differentiation, primordial germ cells, germline chimera

Authors:

Chunhai Liu^{1*}, Il-Kuk Chang¹, Kamal A Khazanehdari¹, Shruti Thomas¹, Preetha Varghese¹, Vijaya Baskar¹, Razan Alkhatib¹, Wenhai Li¹, Jörg Kinne¹, Michael McGrew², Ulrich Wernery¹

¹*Central Veterinary Research Laboratory (CVRL), P.O. Box 597, Dubai, UAE*

²*Division of Developmental Biology, Easter Bush Campus, University of Edinburgh, UK*

*To whom correspondence should be addressed: avian@cvrl.ae

Telephone: +971-4-337 5165, Fax: +971-4-336 8638

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ABSTRACT

Cloning (SCNT) in avian species has proven unachievable due to the physical structure of the avian oocyte. Here, the sexual differentiation of primordial germ cells with genetic sex ZZ (ZZ PGCs,) was investigated in female germline chimeric chicken hosts with the aim to produce uniparental offspring. ZZ PGCs were expanded in culture and transplanted into the same and opposite sex chicken embryos which were partially sterilized using irradiation. All tested chimeric roosters (ZZ/ZZ) showed germline transmission with transmission rates of 3.2-91.4%. Unexpectedly, functional oogenesis of chicken ZZ PGCs was found in three chimeric hens, resulting in a transmission rate of 2.3-27.8%. Matings was conducted between the germline chimeras (ZZ/ZZ and ZZ/ZW) which derived from the same ZZ PGCs line. Paternal uniparental chicken offspring were obtained with a transmission rate up to 28.4% and as expected, all uniparental offspring were phenotypic male (ZZ). Genotype analysis of uniparental offsprings was performed using 13 microsatellite markers. The genotype profile showed that uniparental offspring were 100% genetically identical to the donor ZZ PGC line, shared 69.2-88.5% identity with the donor bird. Homozygosity of the tested birds varied from 61.5-84.6%, which was higher than the donor bird (38.5%). These results demonstrate that male avian ZZ PGCs can differentiate into functional ova in an ovary, and uniparental avian clones are possible. This technology suggests novel approaches for generating genetically similar flocks of birds and for the conservation of avian genetic resources.

INTRODUCTION

In many species, sex determination and sexual differentiation into sex-related phenotypes are events of genetic programming. The genetic gender is determined at the moment of fertilization. In birds, the male possesses the homogametic sex (ZZ), and the heterogametic sex is the female (ZW) [1]. The early precursors of the gametes, the primordial germ cells (PGCs), are formed early in development and migrate to colonize the forming gonad. Germ cell differentiation is guided by the embryonic milieu in many species. Under influences of sex hormones and the surrounding somatic cells, male germ cells in the testes differentiate into the resident spermatogonia and form spermatozoa, while in females the germ cells enter meiosis and differentiate into the ova. PGCs are the progenitor cells of germ cell lineage. Several key pluripotency genes (such as *Oct4*, *Sox2*, *Nanog* etc) are expressed in PGCs [2], and PGCs from various stages of early embryo showed plasticity in their developmental fate in vitro and in vivo. In many vertebrate species, the germ cells will follow the somatic environment and form functional gametes when transplanted into the opposite sex. In the XX testis of sex reversed mouse, XX germ cells enter first phase of spermatogenesis and become prospermatogonia. In the XY ovary, XY germ cells enter meiosis and differentiate as primary oocytes, the fertility depends on the species, genetic background and causes of sex reversal [3]. In fish, ZZ germ cells will form functional ova when transplanted into female host embryos [4]. In birds, the situation is less certain. When blastoderm cells containing precursors of PGCs [5], migratory PGCs [6] and gonadal PGCs [7, 8] were transferred into germ cell-ablated, reverse sex hosts, offspring from the transplanted PGCs were reported at extremely low frequencies. In those studies, the number and purity of donor PGCs were limited. In a defined culture system, chicken PGCs could keep proliferating for a long period and reprogrammed into a germline competent cell line [9]. A high

rate of germline transmission was obtained by transferring cultured PGCs. It was reported that cultured PGCs did not form functional gametes in sex reversed hosts [10]. In present study, Barred Plymouth Rock chicken ZZ PGCs were cultured and transplanted into both male and female irradiated White Leghorn chicken hosts. The oogenesis of chicken ZZ PGCs in the ovary of chimeric chicken was investigated. Donor derived offspring were produced from both sexes. Furthermore, paternal uniparental chicken offspring were produced from the functional spermatozoa and ova of sole genetic origin through male and female germline chimeras. Uniparental chicken offspring may provide desired experimental materials for avian gametogenesis research, and also novel approaches for breeding and conservation of avian genetic resources. (The experimental design was illustrated in Figure 1).

MATERIALS AND METHODS

Barred Plymouth Rock (homozygous recessive i/i) chickens, White Leghorn chicken (homozygous dominant I/I) layers and chimeric birds were kept in the chicken house at the Central Veterinary Research Laboratory (CVRL), Dubai, UAE. All experimental animals and treatment in this study were reviewed and approved by the Animal Ethic Committee of CVRL, and Ministry of Climate change and Environment of the UAE, according to the Ministerial Decree No. 384 of the year 2008 on the executive by-law of the Federal Law No. 16 of the year 2007 concerning animal welfare.

Culture and characterization of Barred Plymouth Rock chicken PGCs

Barred Plymouth Rock chicken PGCs were cultured as described by van de Lavoie *et al* [9]. Briefly, 2-3µl blood samples were collected from 123 Barred Plymouth Rock chicken embryos (stage 14, H&H). The embryos were sealed with double layers of parafilm, and returned to incubator. Donor embryos from which optimal PGCs culture derived were allowed to hatch.

Blood samples were seeded on mitotically inactivated mouse fibroblast feeders (STO, ATCC CRL-1503) with Knockout Dulbecco's modified Eagle's medium (Invitrogen) supplemented with 50% Buffalo Rat Liver cells (BRL 3A, ATCC CRL-1442) conditioned medium, 7.5% fetal bovine serum, 2.5% chicken serum, 1x non-essential amino acid, 1x nucleosides, 10mM sodium pyruvate and β -mercaptoethanol. Fibroblast growth factor (FGF2) and stem cell factor (SCF) were added at a concentration of 10ng/ml each. The cultures were maintained in 5% CO₂ incubator at 37 °C. Cells were passaged at 3-4 days interval. A density of 5×10^4 cells were seeded in each well of 24 well plate (Falcon, Corning USA).

PGCs were fixed in 4% paraformaldehyde, permeabilized in 0.5% Triton X-100, and incubated in blocking buffer (PBS/5%BSA/0.5%Tween-20) for 1 hour. Anti-SSEA-1 (stage-specific embryonic antigen 1, MC480, 1:200) was added and incubated at 4°C overnight, then with secondary antibody biotinylated goat anti-mouse IgM (BA2020, Vector laboratories) for 1 hour. Texas Red Avidin D (A-1100, Vector laboratories) was added at 1:500 for 1 hour, washed with PBS (-) and mounted with Vectashield/DAPI. Slides were observed under fluorescent microscope. Chicken VASA homologue were also detected with the same procedure, except using rabbit anti-CVH (1:500, a gift from Dr. Noce, Keio University, Japan) and rabbit polyclonal anti-DDX4/MVH (mouse VASA homologue, Abcam ab13840) as primary antibody, and goat anti-rabbit biotinylated (1:200) as secondary antibody. PGCs omitting primary antibody incubation were used as negative control. The experiment was repeated twice.

To detect gene expression profiles of chicken PGCs, total RNA was extracted using the RNeasy mini kit (Qiagen). RT premix was prepared as the manufacturer's instruction. 15 μ l premix was added to 10 μ l of total RNA (1 μ g). cDNA was obtained using Ipsogen RT kit (Qiagen). The PCR primers for detecting the expression of *cPouV*, *cNanog*, *cKlf4*, *cSox2*, *cMyc*, *Cvh* and *Gapdh*

genes were applied as published [10]. Primers detecting *cDazl* gene was designed: forward 5'AGAAGGAAAAATCATGCCAAA, reverse 5'ATTGCTGGTCCCAGTTTCAG. PCR was performed using Taq PCR master mix kits (Qiagen). Reaction mixture was prepared as 12.5µl master mix, 1.25µl forward primer, 1.25µl reverse primer, 8µl H₂O, 2µl cDNA. PCR was run at 95°C, 20 minutes; 95°C, 30 seconds; 55-59°C, 30 seconds; 72°C, 1minute; 60°C for 30 minutes. PCR products were loaded on 1% agarose gel.

Transplantation of Barred Plymouth Rock chicken ZZ PGCs into White Leghorn chicken embryos

Freshly laid White Leghorn chicken eggs were collected, and irradiated with an X-ray facility at a dose of 600 rads in Al Tawam hospital, Al Ain, UAE. The non-irradiated normal eggs were used as control. All the recipient eggs were incubated to stage 16-17 (H&H). To expose the embryo, a small piece of egg shell was removed from the sharp end of eggshell. Barred Plymouth Rock chicken ZZ PGCs in culture were harvested by gently pipetting, centrifuged and resuspended to a concentration of 2×10^3 cells/µl in the culture medium. A number of 4,000 PGCs in 2 µl culture medium were injected into the dorsal aorta of recipient White Leghorn chicken embryos [11]. Two ZZ PGCs lines (1110g.64, 1110h.79) were selected as donor cells to produce germline chimera. The injected eggs were sealed with double layer of parafilm, and incubated at 37.8 °C with 60% relative humidity to hatch.

Progeny test of chimeric chickens

The chimeric birds were raised to sexual maturity under common condition. To evaluate the contribution of donor PGCs in the germline of recipient birds, 28 putative male chimeras and 27 female chimeras (ZZ/ZW) were selected for progeny test through artificial insemination. The

resulting black chicks with Barred Plymouth Rock plumage were considered as donor PGCs derived offspring.

Phase 1. Screening male germline chimera (ZZ/ZZ) and female germline chimeric females (ZZ/ZW)

The semen samples were collected individually from 28 putative male chimeras (ZZ/ZZ), and inseminated into normal Barred Plymouth Rock females. 27 chimeric females (ZZ/ZW) were inseminated with normal Barred Plymouth Rock chicken semen. Insemination was conducted once a week for three months. The resulting eggs were collected and incubated to hatch.

Phase 2. Generation of uniparental Barred Plymouth Rock chicken offspring.

Two male germline chimeras (ZZ/ZZ) and 3 female chimeras (ZZ/ZW) showed high rate of germline transmission, and were crossbred through artificial insemination. The resulting eggs were incubated to hatch. The dead embryos during the last week of incubation were opened to check the feather phenotype. Molecular sexing was carried out by PCR with feather samples from the resulting chicks with Barred Plymouth Rock phenotype. These chicks were raised to sexual maturity under common condition, and bred with normal Barred Plymouth Rock females to confirm the fertility.

Molecular analysis of paternal uniparental chicken offspring

The genotype analysis of genomic DNA was carried out using 13 chicken microsatellite markers (supplementary data: table S3) [12, 13, 14]. The 5' end of the forward primer was labeled with a universal M13 forward or M13 reverse tail. Fluorescent labeled M13 primers were used to detect amplified products. The primers for various loci were multiplexed in the way that the amplified product sizes did not overlap. Genomic DNA of uniparental offsprings, surrogate parents, the donor bird and PGCs line was extracted using the Qiagen DNeasy blood and tissue extraction kit.

Extracted DNA was checked using the Nanodrop (Thermo Scientific), and diluted to a concentration of 30-50 ng/μl. The PCR was performed in a total volume of 11 μl mixture containing 30-50 ng genomic DNA, 2 mM MgCl₂, 200μM of each dNTP, 1U GoTaq polymerase (Promega Inc. WI). The PCR cycling conditions were as follows: 95°C for 3 minutes followed by 35 cycles of 95°C for 15sec, 54°C for 30 sec, 72°C for 30 sec followed by final extension at 72°C for 45 minutes and 60°C for 30 minutes. Normal chicken genomic DNA was used as positive control, and non-template PCR mix as negative control. Amplified PCR products were mixed with Hi-Di formamide, internal size standard GeneScan-Liz 500 (Applied Biosystems, USA), denatured at 95°C, and snap cooled. Size analyses of the separated DNA fragments were performed using the GeneMapper V4 software (Applied Biosystems, USA).

Statistical Analysis

The proportion data of germline transmission frequencies were Arcsine transformation, then analyzed with Student's *t*-test. The hatchability differences were subjected to *Chi*-Square tests. Statistical Analysis was performed on SPSS software version 20 (Statistical Analysis System, SAS Institute, 1990). Statistical significance was accepted at $P < 0.05$.

RESULTS

Derivation of Barred Plymouth Rock chicken primordial germ cell line from embryonic blood samples.

A total of 123 blood samples (59 male, 64 female) were collected and cultured. PGCs divided slowly in the first a few days, and were seen after a week. Cells attached loosely on the STO feeder layer, and were harvested by gentle pipetting. After 20-30 days culture, a total number of five ZZ PGC cultures grew robustly (Figure 2 a) with a population doubling time about 20 hours, were maintained for transplantation and frozen down. ZW PGCs showed different growth

pattern, proliferated slower than ZZ PGCs, and aggregated into tightly compacted clumps floating on the feeder layer. In present study, two ZW PGC lines were obtained from 64 primary cultures. Out of the 59 male embryos, 15 chicks hatched with a hatchability of 12.2%. Two ZZ PGC lines (1110g.64 and 1110h.79) were selected and cultured for 38-49 days for chimera production.

Cultured chicken PGCs were stained positively with the antibody recognizing SSEA-1 and anti-CVH, weakly stained with anti-DDX4/MVH antibody. No signal was shown with the control cells when omitting primary antibody (Figure 2 c). Pluripotency-associated genes (*cPOUV*, *cNANOG*, *cSOX2*), oncogenic genes (*cMYC* and *cKLF4*), and germ cell specifically expressed genes (*DAZL*, *CVH*) were detected from the two ZZ PGC lines. PCR products were amplified from the cDNA samples with all 8 set primers. These results in accordance with literatures of other researchers [9,10], suggested that cultured PGCs expressed *POUV*, *NANOG*, *KLF4*, *SOX2* and *c-MYC* homologous genes, and the expression of *DAZL*, *VASA* confirmed their germ cell lineage identity (Figure 2 b).

Production of male (ZZ/ZZ) and female (ZZ/ZW) chicken germline chimeras

A total number of 214 white leghorn chicken eggs were injected with chicken ZZ PGC lines (1110g.64 and 1110h.79), and 80 eggs hatched with the hatchability of 37.4%. Among the injected eggs, 34.5% (38/110) of the irradiated eggs hatched, and 40.4% (42/104) of the eggs hatched from the non- irradiated control eggs. There was no significant loss of hatchability with X-ray irradiation (Chi-Square test, $P > 0.05$). A total number of 80 chimeric chicks were raised under normal condition. Seventy birds (38 male chimeras and 32 ZZ/ZW type female chimeras) survived at 2 months old (Table 1).

Germline transmission of male (ZZ/ZZ) chimeric chicken

Twenty eight (100.0%) birds produced donor PGCs derived black Barred Plymouth Rock chicks in the male chimeras (ZZ/ZZ), and were confirmed as germline chimera. Donor PGC line 1110g.64 derived Barred Plymouth Rock chicken progenies was $54.1 \pm 23.3\%$ in non- irradiated recipient eggs and $81.7 \pm 10.1\%$ in irradiated recipients (Table 2). In the chimeric roosters (ZZ/ZZ) from PGC line 1110h.79, the proportion of Barred Plymouth Rock chicken offspring was $53.3 \pm 24.3\%$ in non-irradiated recipient eggs, and $54.5 \pm 12.5\%$ in irradiated recipients (supplementary data, table S1). Prior to introducing donor PGCs, irradiation of recipient embryos significantly increased the germline transmission rate in chimeras with cell line 1110g.64 (Student's *t*-test, $P < 0.05$); however, there was no significant difference in the birds transplanted with PGC line 1110h.79.

Germline transmission of opposite-sex chimeric females (ZZ/ZW)

Among 11 chimeric females (ZZ/ZW) from the PGCs line 1110g.64, three birds (no.17, x103, x126) produced donor derived Barred Plymouth Rock chicken offsprings with the transmission rate of 2.3%, 2.6% and 27.8% (Table 3, Figure 2 d, e, f), respectively. All of the black offsprings obtained from female chimeras (ZZ/ZW) were identified as males (ZZ) by molecular sexing with PCR (Figure 2 g). However, the 16 chimeric females (ZZ/ZW) from the PGC line 1110h.79 did not produce any Barred Plymouth Rock chicken offsprings (supplementary data, Table S2).

Generation of uniparental Barred Plymouth Rock chicken offsprings

Three chimeric females (ZZ/ZW; bird no.17, x103 and x126) and two chimeric roosters (ZZ/ZZ; bird x104, x143) which showed high germline transmission rate in the progeny test phase I (Table 3,4), were chosen to participate the testcrossing in phase 2 with the aim to produce uniparental offspring. The combinations of these birds were set as follows (Table 4):

A total number of 20 offsprings with Barred Plymouth Rock chicken phenotype were obtained from the 2 chimeric females (ZZ/ZW). The bird x126 showed 28.4% (19/67) of germline transmission rate. These Barred Plymouth Rock offsprings were paternal uniparental offspring derived from the solo genetic source cell line 1110g.64. Molecular sexing results showed that all of the 20 Barred Plymouth Rock offsprings were male (ZZ). A number of 6 offsprings died during the third week of incubation, and 7 chicks died in the first week after hatch, 7 survived to sexual maturity. The paternal uniparental offsprings showed high mortality which could be caused by high extent of homozygous genotype.

Molecular analysis of paternal uniparental Barred Plymouth Rock chicken offsprings

Thirteen individual microsatellite primer pairs were used (Table 5). Nine PCR products were found specific to genomic DNA of donor PGCs line, with the sizes of 190bp (MCW14), 336bp (MCW183), 153bp (MCW81), 133bp (HUI12), 144bp (HUI2), 165bp (MCW216), 120bp (MCW295), 245bp (MCW34), and 173bp (MCW69). Particularly, three primer pairs (MCW14, MCW183 and MCW81) showed homozygous loci with the genomic DNA of donor PGCs line, and produced specific PCR product size of 190bp, 336bp and 153bp respectively. With the 13 primer pairs, genotype of the PGC line (1110g.64) was 100.0% identical to the donor Barred Plymouth Rock rooster (BPR64). Uniparental offsprings showed only donor specific PCR products in the three homozygous loci with primer pairs (MCW14, MCW183 and MCW81). In addition, all of the uniparental offsprings showed the Barred Plymouth Rock (homozygous recessive i/i) plumage phenotype. These genotyping results confirmed that uniparental offsprings genetically originated from the donor PGCs line (1110g.64) only, and were non-related to the surrogate parents (White Leghorn chimeric chicken x104 and x126) (supplementary Figure S1). Among the 26 alleles of 13 loci, 18 to 23 alleles of uniparental offsprings were identical to the

donor bird (BPR64), indicating that the genome of uniparental offsprings were 69.2 % (18/26) to 88.5% (23/26) identical to that of donor bird (BPR64). Among the 13 loci of genomic DNA in donor bird, 5 (38.5%) loci (MCW111, MCW14, MCW183, MCW284, and MCW81) showed homozygous. However, in the 6 uniparental offsprings tested, 8 to 11 loci were found homozygous, indicating the homozygosity of 61.5% (8/13) to 84.6% (11/13), which was much higher than that of the donor bird (38.5%). These results suggested that uniparental offsprings bred through “self-mating” inherited higher genetic content of the only uniparent, and greatly improved the homozygosity than conventional breeding.

DISCUSSION

Primordial germ cells are the progenitor cells of germline development. In birds, precursors of PGCs were firstly identified as about 30 cells expressing *Vasa* homologue gene (*Cvh*) in the central zone of area pellucida at stage X [15]. In mammal, PGCs proliferated in vitro and reprogramed into embryonic germ cells (EG) in a medium with the addition of several growth factors [16, 17]. However, in similar culture conditions, chicken PGCs proliferate for long-term, and are still capable to repopulate in the gonad of recipient birds, giving rise to donor derived offspring [9,10]. In the present study, a number of 5 (8.5%) ZZ PGC lines were established from 59 cultures of male samples; and 2 (3.1%) ZW PGC lines were obtained from 64 cultures of female samples. In general, ZZ PGCs proliferate faster than ZW PGCs in primary cultures, but also individual variation was found among the cultures of both sexes [18]. In most cases, ZW PGCs divide a couple of times, form tightly compacted clumps in the early passages and proliferate slowly. Chicken PGCs were found expressing pluripotency genes (such as *Oct-4*, *Sox2*, *Nanog* etc.) at early stage. The transcription of these genes was gradually down-regulated from stage 14 to 28 [2]. As a consequence, more PGCs might progressively lose the pluripotency

in advanced developmental stages. The proliferation discrepancy of PGCs in culture may attribute to their developmental heterogeneity. In similar studies, Song et al 2014 reported that the culture of PGCs has a higher rate of success from germinal crescent area than those from blood source and primitive gonadal tissues [18]. Long-term cultured PGCs were reported to keep expressing pluripotency genes and germline specific genes. Our present study also confirmed the expression of *cPouv*, *cNanog*, *cKlf4*, *cSox2*, *cMyc*, *cDazl* and *Cvh* genes.

The transmission rate of chimera was reported to improve by altering the ratio of donor and recipient germ cells by irradiation [19], chemical treatment [20], or physical removal [5]. In our present study, the recipient eggs were irradiated with X-ray, which reduced up to 94.8% of endogenous germ cells at stage 28 (data not shown). In the present study, X-ray irradiation significantly improved the transmission rate from 54.1% to 81.7% in the group with cell line 1110g.64. However, irradiation did not significantly improve the transmission rates of the chimeras from the cell line 1110h.79. This may be possibly caused by a different batch of irradiation.

Many reports have indicated that the sex differentiation of PGCs is directed by the external signals in the gonadal environment, and not by the germ cells themselves [21]. Differentiation of chicken ZZ PGCs in the opposite-sex female chimera were reported by transferring circulating, gonadal PGCs and blastoderm cells. The germline transmission rate was only 0.4-0.9% in the chimera (ZZ/ZW) by transferring circulating PGCs [6], and gonadal PGCs [8]. The extremely low transmission rate made the result inconclusive. In the present study, when transferring large number of pure ZZ PGCs into irradiated recipient male embryos, the frequency of germline transmission were 34.1 times higher than that in female chimeras (81.9% vs 2.4%). Particularly, in the bird x126, ZZ PGCs derived functional ova contributed 27.8% (5/18) of the offspring.

These results suggest that increasing the ratio of exogenous germ cells by transferring higher number of in vitro cultured PGCs and deletion of endogenous PGCs can also improve the chances of ZZ PGCs in functional ova production.

The plasticity of sex differentiation in germ cell has also been reported in other species. For instance, germ cells of rainbow trout at various developmental stages, eg PGCs, spermatogonia stem cells, and pre-meiotic oogonia, were capable to colonize in sexually undifferentiated embryonic gonad of the same or opposite sex, and further gave rise to both, functional spermatozoa and eggs [4,22]. Chicken testicular and ovarian gonocytes from 20 day-old embryos were still capable to migrate, and contribute to germline after transfer into recipient embryos at stage 14 [23]. It is likely that the plasticity of sex differentiation of germ cell remained until spermatogonia stem cells in male and pre-meiotic oogonia in female. However, it is not clear whether these developmentally advanced germ cells will adapt to the pace with endogenous germ cells or maintain the existing developmental process at their own pace. The germline transmission of opposite-sex chimera confirmed the possibility of functional ova differentiation from chicken ZZ PGCs. However, the efficiency of ZZ PGCs differentiation into ova is still far lower than that into spermatozoa in nature. The generation of functional ova from PGCs is a long process involving migration, colonization, mitotic proliferation, differentiation and meiotic stages. When transferring equal number of chicken gonad PGCs, Nakajima et al 2014 reported that the number of donor PGCs did not show significant difference in the gonad of recipient embryos among any of the 4 possible same and opposite sex combinations [7]. In the model of sex reversed male chicken (ZW), Albinawanto et al [24] found that the ZW germ cells could complete the second meiosis, and the transformation of spermatid to spermatozoa was partially impaired. It is still unknown how and at what stage of oogenesis ZZ PGCs differ from their

counterpart the endogenous ZW PGCs, either during mitotic proliferation or the process of meiosis in the ovary of the host.

What made us curious was to cross the female (ZZ/ZW) and male (ZZ/ZZ) chimeras of the same donor PGCs origin. Compared to conventional inbreeding program, the uniparental offspring were expected to preserve or restore most of the genetic information of a specific donor rooster. In the present study, genotyping results showed that the homozygosity of the 13 loci from the detected 6 offsprings varied from 61.5 to 84.6% (Table 5), which increased 30-40% than that of the donor birds (38.5%) in one generation. In addition, the comparison of genotype profile in the 13 loci showed that the uniparental offspring was 100% derived donor PGCs. In present method, the donor rooster transmitted 69.2-88.5% genetic information to his uniparental offspring, instead of 50% in conventional breeding. Breeding using opposite-sex chimera will enhance the chances to achieve higher homozygosity in a shorter period than a conventional inbreeding program, and provide a new approach to generate specific chicken line with interesting traits in a short time period. The complete oogenesis of ZZ PGCs may reveal that W chromosome is unessential for ova development. Additionally, in sexual reproduction or somatic cell nuclear transfer (SCNT) derived animals, the mitochondrial DNA is maternally inherited. Although the mitochondrial DNA information of these uniparental chicken clones was not analyzed, it likely originated from the donor PGCs. Therefore, uniparental chickens may provide a desired model for gametogenesis research, and novel approaches for conservation and restoration of the avian genetic resources.

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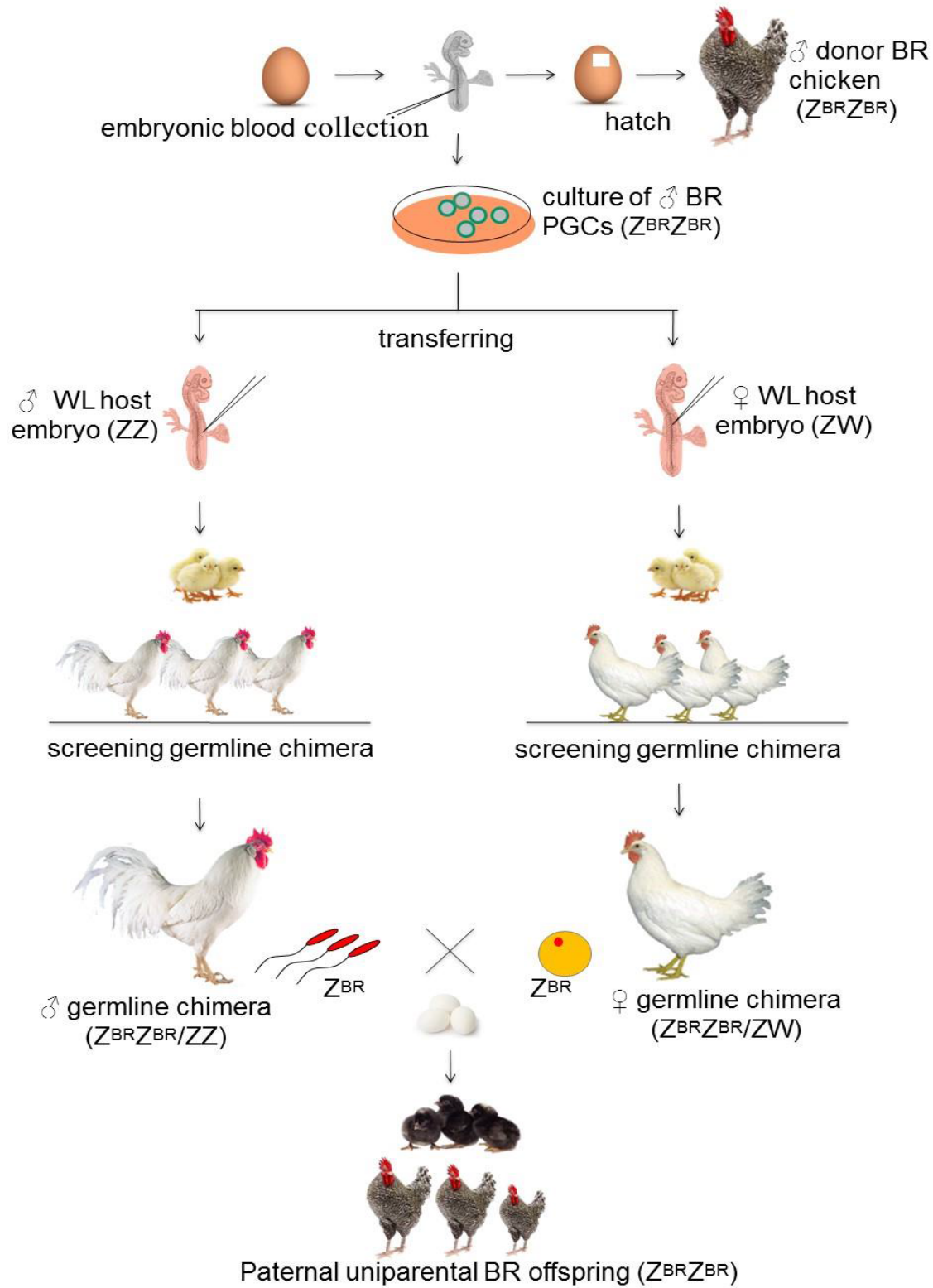


Figure 1. Diagram of experimental design and procedures

Figure 2. Culture, characterization of chicken PGCs, and generation of paternal uniparental offspring. (scale=20µm)

- (a) Barred Plymouth Rock chicken PGCs (arrow, 1110g.64) 38 days in culture.
- (b) gene expression profile of cultured chicken PGCs (*Cvh*, *cDazl*, *cPouv*, *cNanog*, *cSox2*, *cKlf4* and *cMyc*).
- (c) characterization of chicken PGCs with anti-SSEA-1, anti-CVH and anti-MVH.
- (d) newly hatched paternal uniparental offspring.
- (e) a dead uniparental embryo .
- (f) heterosexual (ZZ/ZW) chimeric hen (x126), germline chimeric rooster (x104) and their paternal uniparental offspring.
- (g) molecular sexing of uniparental offspring.

Table 1 Transplantation of cultured Barred Plymouth Rock chicken PGCs (ZZ) into White Leghorn chicken embryo

group	recipient WL eggs	donor BR PGCs (ZZ) line	chimera production			survival at 2 month-old		
			No. eggs injected	No. eggs hatched	hatchability	male ZZ/ZZ	female ZZ/ZW	total
1	normal WL	1110g.64	45	19	42.2%	9	7	16
2	irradiated	1110g.64	55	20	36.4%	11	7	18
3	normal WL	1110h.79	59	23	39.0%	12	8	20
4	irradiated	1110h.79	55	18	32.7%	6	10	16

Table 2 Progeny test of male chimeras (ZZ/ZZ) from PGC line 1110g.64

group	bird ID	No. of white offspring	No. of black offspring	total No. of offspring	rate of germline transmission
group 1 normal recipient	2	10	51	61	83.6%
	3	46	16	62	25.8%
	36	15	47	62	75.8%
	39	9	5	14	35.7%
	40	10	7	17	41.2%
	44	32	53	85	62.4%
	means \pm SD				54.1 \pm 23.3%
group 2 irradiated recipient	x104	5	43	48	89.6%
	x108	5	26	31	83.9%
	x116	3	10	13	76.9%
	x129	18	55	73	75.3%
	x140	3	32	35	91.4%
	x142	10	18	28	64.3%
	x143	3	29	32	90.6%
	means \pm SD				81.7 \pm 10.1%*

(*Student's *t*-test, $P < 0.05$)

Table 3 Progeny test of opposite-sex chimeric females (ZZ/ZW) with PGC line 1110g.64

group	bird ID	No. of white offspring	No. of black offspring	total No. of offspring	rate of germline transmission
group 1 normal recipient	5	38	0	38	0
	17	42	1	43	2.3%
	21	48	0	48	0
	27	23	0	23	0
	42	38	0	38	0
	total	189	1	190	0.5%
group 2 irradiated recipient	x103	38	1	39	2.6%
	x125	61	0	61	0
	x126	13	5	18	27.8%
	x132	58	0	58	0
	x150	30	0	30	0
	x152	44	0	44	0
	total	244	6	250	2.4%

Table 4 Testcross between male (*ZZ/ZZ*) and female (*ZZ/ZW*) chimeras from the same PGC line
(1110g.64)

combination ♂-♀	No. of eggs	fertility	No. of offspring			incubation and survival of black offspring			
			white offspring	black offspring	% of black offspring	< 13dpi*	14- 21dpi	died 1 st week	adult
x104-x126	83	81.9%	48	19	28.4%	N/D	6	7	6
x143-x103	74	32.4%	23	1	4.2%	N/D	0	0	1
x143-no.17	76	55.3%	42	0	0	N/D	0	0	0

(dpi: days post incubation)

Table 5 Genotype of paternal uniparental Barred Plymouth Rock offspring from chimeras (ZZ/ZZ and ZZ/ZW)

marker	genetic parent		genotype of uniparental Barred Plymouth Rock offspring						surrogate parents			
	Donor bird BPR64	Donor cell line 1110g.64	x126-1	x126-2	x126-3	x126-4	x126-5	x126-6	chimera (ZZ/ZZ) x104	chimera (ZZ/ZW) x126		
MCW111	117/117	117/117	117/117	117/117	117/117	117/117	117/117	117/117	117/117	117/115		
MCW14	190/190	190/190	190/190	190/190	190/190	190/190	190/190	190/190	204/206	204/206		
MCW183	336/336	336/336	336/336	336/336	336/336	336/336	336/336	336/336	312/312	324/320		
MCW284	263/263	263/263	263/263	263/263	263/263	263/263	263/263	263/263	263/263	263/263		
MCW81	153/153	153/153	153/153	153/153	153/153	153/153	153/153	153/153	131/131	131/131		
HUJ12	133/153	133/153	133/153	153/153	133/133	133/153	153/153	133/153	137/153	135/153		
HUJ2	144/158	144/158	144/144	144/158	158/158	144/144	144/144	144/158	148/158	148/150		
MCW216	165/167	165/167	165/167	165/165	165/167	167/167	167/167	165/167	167/167	167/167		
MCW295	106/ 120	106/120	106/120	106/120	106/120	106/106	106/120	120/120	104/102	106/104		
MCW330	284/292	284/292	284/284	284/292	292/292	284/284	ND	284/292	284/292	292/292		
MCW34	241/ 245	241/245	241/245	245/245	241/245	245/245	ND	241/241	241/249	251/253		
MCW67	195/199	195/199	195/195	195/199	199/199	195/199	195/ND	195/195	195/199	195/195		
MCW69	173/179	173/179	173/173	173/173	173/179	173/173	173/179	173/179	179/179	179/179		
homozygosity	5/13	5/13	9/13	9/13	9/13	11/13	8/13	8/13	6/13		6/13	
identity	26/26	26/26	22/26	22/26	22/26	20/26	18/26	23/26	-	-	-	-

Note:

1. the numbers stand for the PCR product fragment size by base pair. *ND: not detected due to technical failure.
2. X126-1,2,3,4,5,6: the six paternal uniparental Barred Plymouth Rock chicken offspring from chimeric rooster (x104, ZZ/ZZ) and chimeric female (x126, ZZ/ZW).
3. in donor PGCs line (1110g.64), 5 loci markers (MCW111, MCW14, MCW183, MCW284 and MCW81) show homozygous. The remaining 8 loci are heterozygous.
4. the numbers in bold shows the 8 loci which gave specific size of PCR product in the genomic DNA of donor bird only, and not existed in the chimeras. The shaded numbers are the heterozygous loci which uniparental offspring inherited from the uniparent.